Technical information

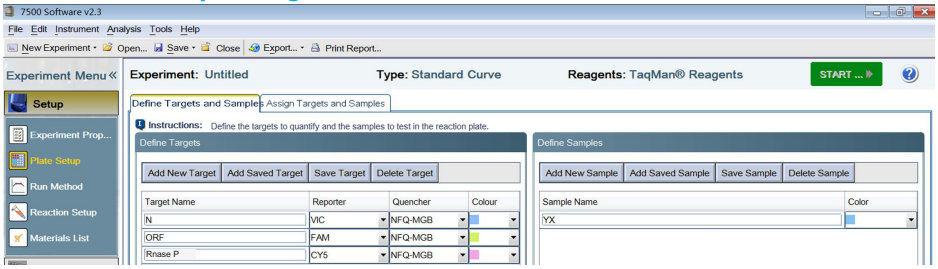
QF 24 V1
CV2 2020

SARS-CoV-2 RT-PCR Detection Kit

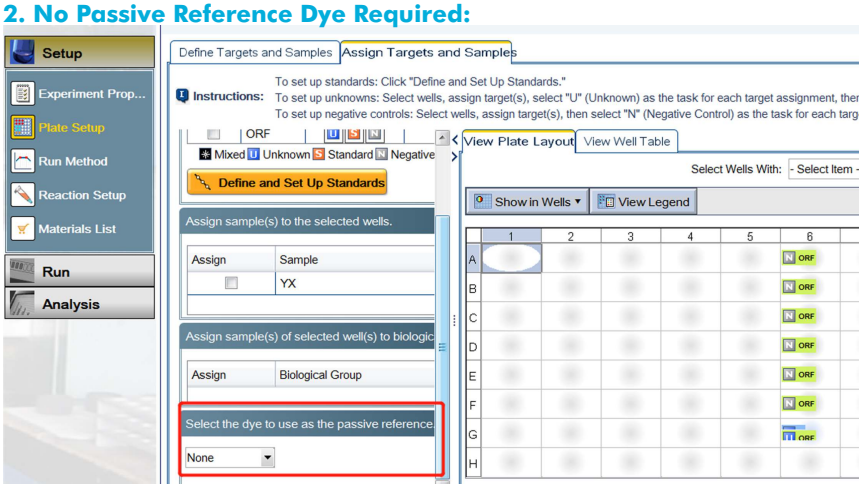
Catalog #: COV-2-RTPCR
Setup for ABI7500: Page 1-2
Setup for BioRad CFX96: Page 2-3

Setup for ABI 7500

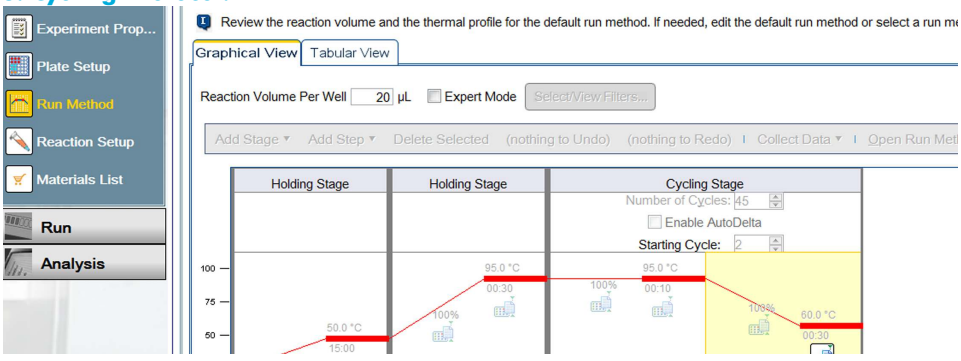
1. ABI 7500 Setup for Signal Collection:



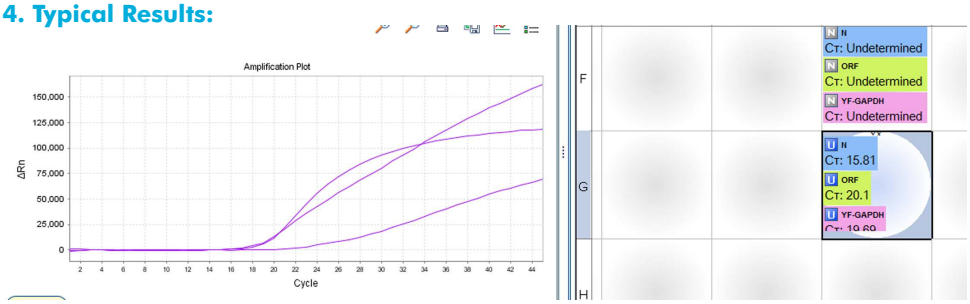
2. No Passive Reference Dye Required:



3. Cycling Protocol:



4. Typical Results:



5. Additional Information:

Do we need to select Rox as a passive reference dye or should that be none?

– No passive reference required.

Do you have DMSO in your reaction mix?

– No.

Why does your kit recommend 45 cycles?

– Some positive samples show at Ct from 37-40, 45 cycles is for sufficient amplication.

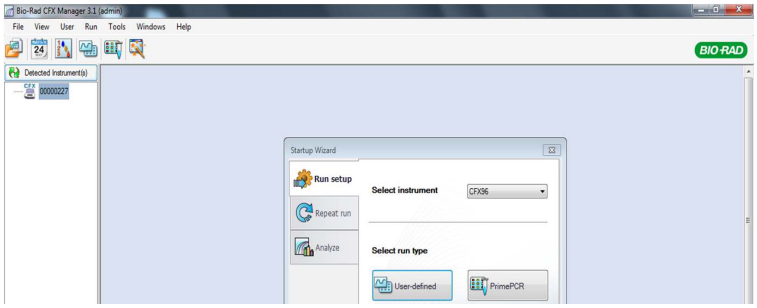
6. Limitations of Use:

Interpretation of results must account for the possibility of false negative and false positive results.

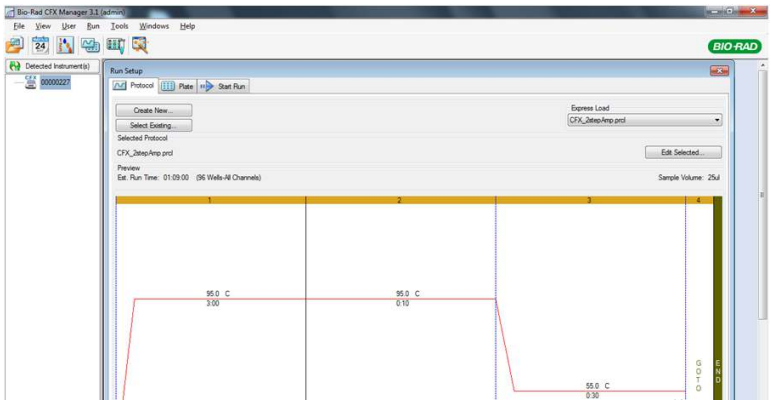
False negatives may be caused by:	False positives may be caused by:
• Unsuitable collection, handling and/or storage of samples. • Sample outside of viraemic phase. • Failure to follow handbook procedures. • Use of unauthorised extraction kit or PCR platform.	• Unsuitable handling of samples containing high concentration of SARS-CoV-2 viral RNA or positive control template • Unsuitable handling of amplified product.

Set up on BioRad CFX96

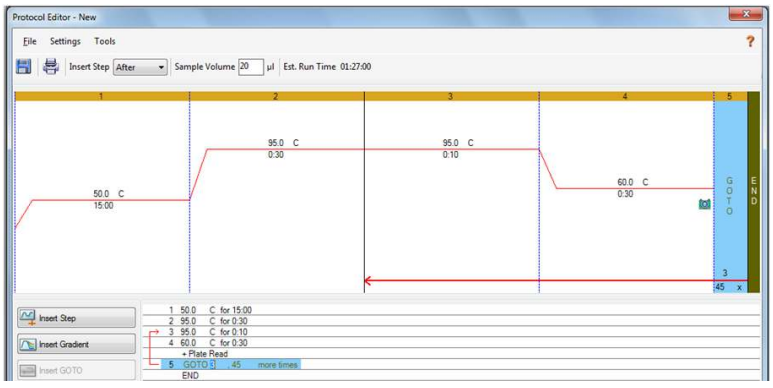
1. Open Bio-Rad CFX Manager Software and select "User-Defined" Run Type.



2. Under Run Setup > Protocol, select "Create New..." to create a new run protocol.

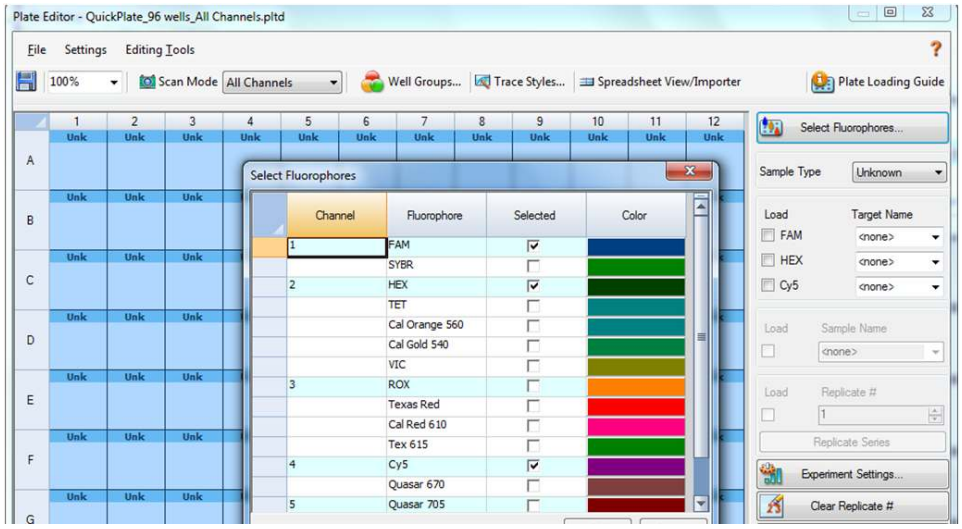


3. Create Cycling Protocol according to the datasheet. Click "OK".

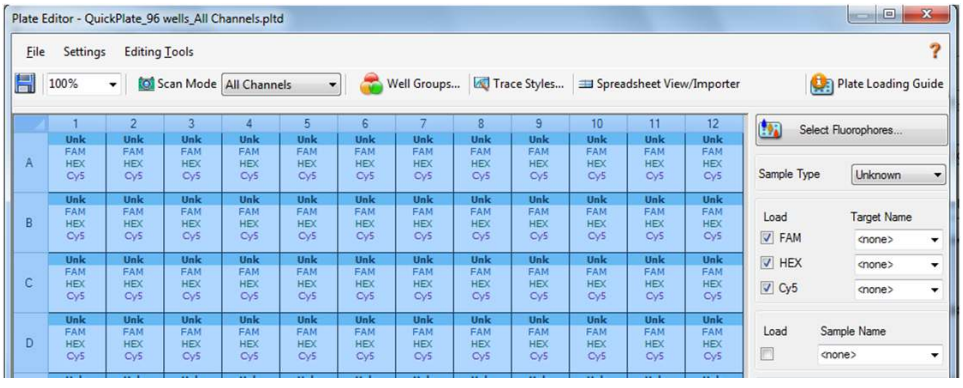


4. Under Run Setup Plate, select "Create New..." to create a new Plate set up.

5. Use the "Select Fluorophores" Option to define channels: HEX (VIC), FAM and CY5. Click "OK".



6. Apply the three channels to all test wells by selecting the entire plate and checking off "Load" FAM, HEX and CY5. Click "OK".



7. Under Run Setup > Start Run, Click on "Start Run" to start the run.