

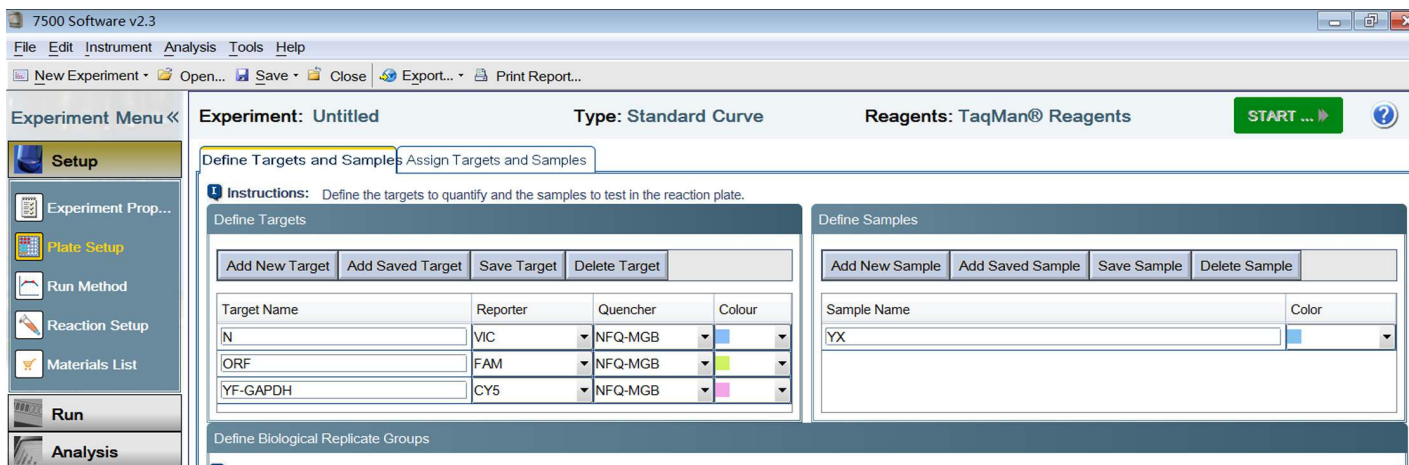
Technical information

QF 24 V1
CV1 2020

SARS-CoV-2 RT-PCR Detection Kit

Catalog #: COV-2-RTPCR

1. ABI 7500 Setup for Signal Collection:



7500 Software v2.3

File Edit Instrument Analysis Tools Help

New Experiment Open... Save Close Export... Print Report...

Experiment Menu << Experiment: Untitled Type: Standard Curve Reagents: TaqMan® Reagents START ... ?

Define Targets and Samples Assign Targets and Samples

Instructions: Define the targets to quantify and the samples to test in the reaction plate.

Define Targets

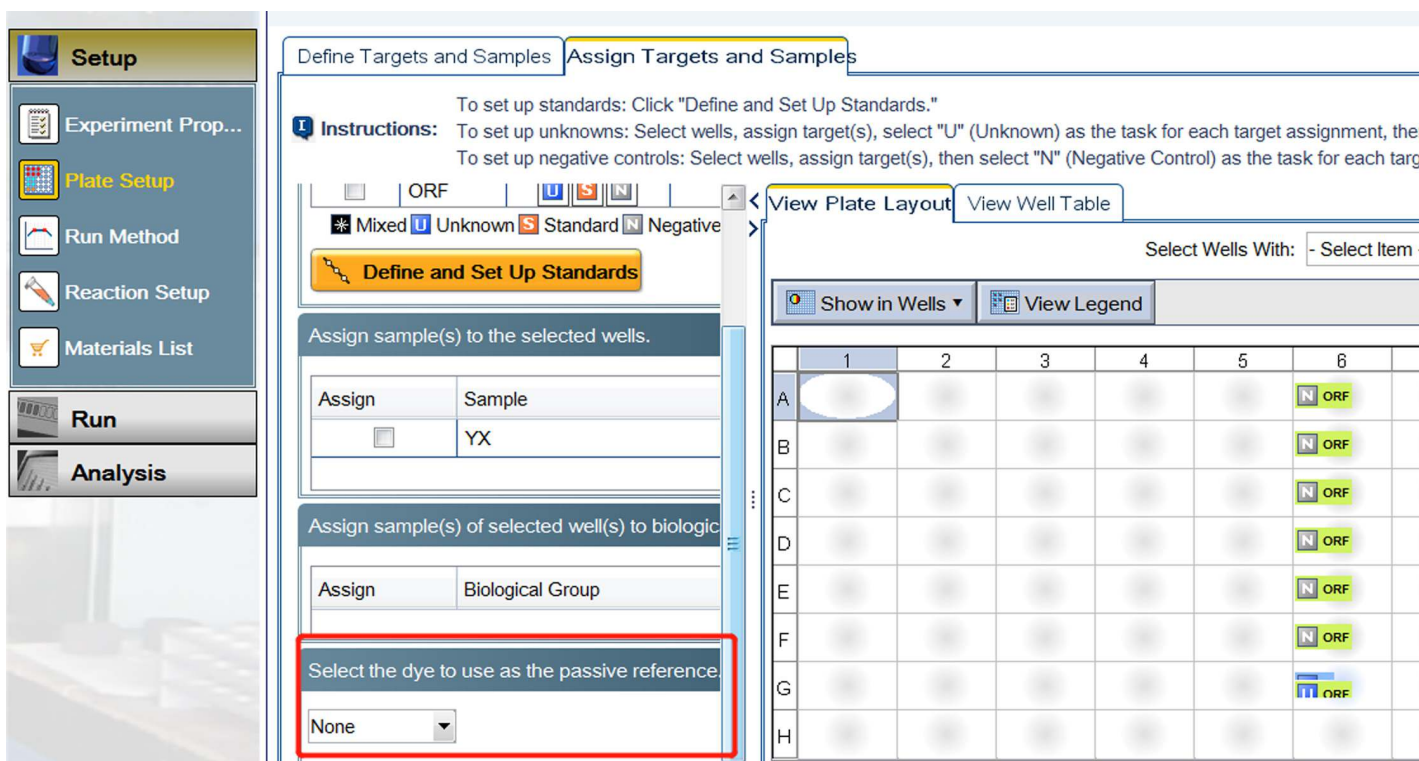
Target Name	Reporter	Quencher	Colour
N	VIC	NFQ-MGB	Blue
ORF	FAM	NFQ-MGB	Green
YF-GAPDH	CY5	NFQ-MGB	Magenta

Define Samples

Sample Name	Color
YX	Blue

Define Biological Replicate Groups

2. No Passive Reference Dye Required:



Define Targets and Samples Assign Targets and Samples

Instructions: To set up standards: Click "Define and Set Up Standards."
To set up unknowns: Select wells, assign target(s), select "U" (Unknown) as the task for each target assignment, then
To set up negative controls: Select wells, assign target(s), then select "N" (Negative Control) as the task for each target assignment.

ORF U S N Mixed U Unknown S Standard N Negative

Define and Set Up Standards

Assign sample(s) to the selected wells.

Assign	Sample
<input type="checkbox"/>	YX

Assign sample(s) of selected well(s) to biological replicate groups.

Assign	Biological Group
<input type="checkbox"/>	

Select the dye to use as the passive reference.

None

View Plate Layout View Well Table

Select Wells With: - Select Item -

Show in Wells View Legend

	1	2	3	4	5	6
A	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/> N ORF
B	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/> N ORF
C	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/> N ORF
D	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/> N ORF
E	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/> N ORF
F	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/> N ORF
G	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/> U ORF
H	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

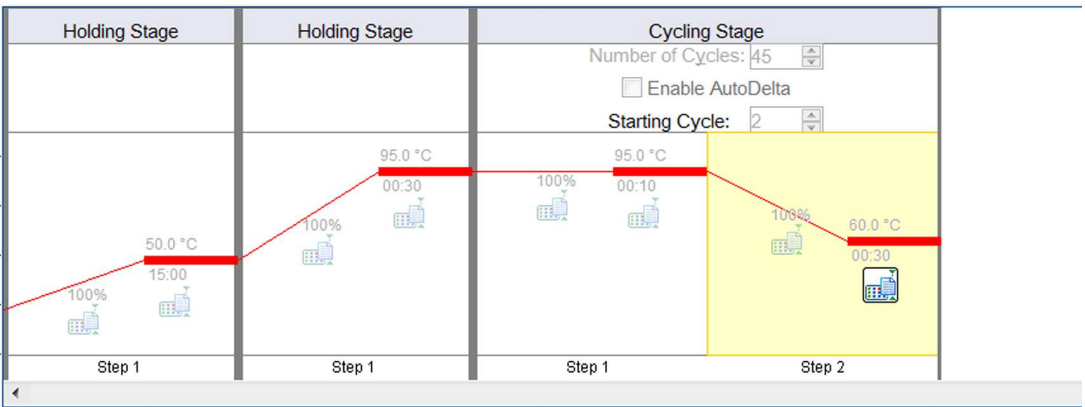
3. Cycling Protocol:

Review the reaction volume and the thermal profile for the default run method. If needed, edit the default run method or select a run method.

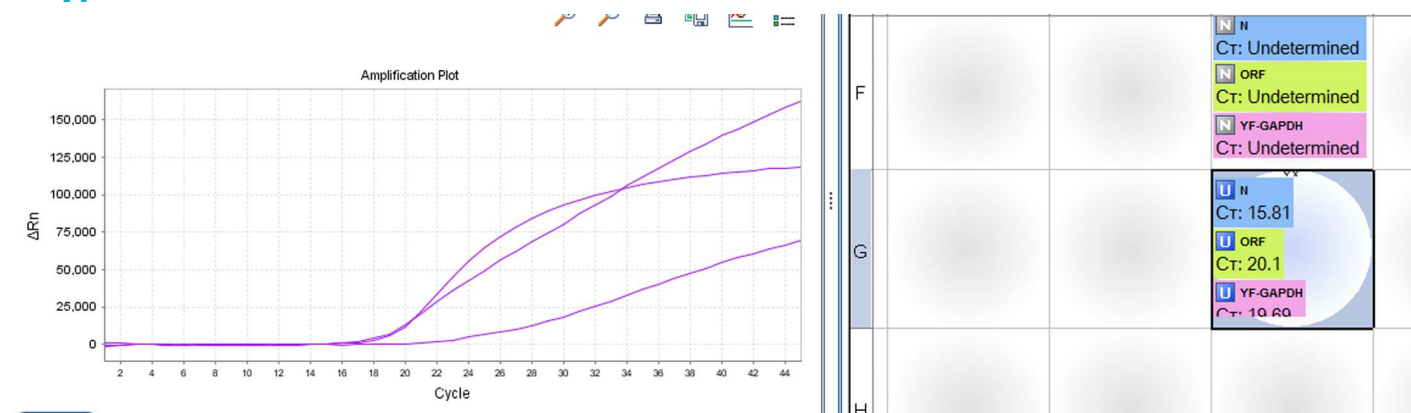
Graphical View | Tabular View

Reaction Volume Per Well: 20 μ L Expert Mode

Add Stage | Add Step | Delete Selected (nothing to Undo) (nothing to Redo) | Collect Data | Open Run Method



4. Typical Results:



5. Additional Information:

Do we need to select Rox as a passive reference dye or should that be none?

- No passive reference required.

Do you have DMSO in your reaction mix?

- No.

Why does your kit recommend 45 cycles?

- Some positive samples show at Ct from 37-40, 45 cycles is for sufficient amplification.